Review Article

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Pseudomonas aeruginosa in Fish and Fish Products: A Review on the Incidence, Public Health Significance, Virulence Factors, Antimicrobial Resistance, and Biofilm Formation

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Abstract

Pseudomonas aeruginosa (*P. aeruginosa*) is a psychotropic pathogenic bacterium that is considered one of the most common spoilage microorganisms related to seafood's consumption. *P. aeruginosa* is widely distributed in nature and isolated from soil, plants, animals, and water. Because of its high resistance to a wide range of antibiotics, *P. aeruginosa* is more dangerous than other spoilage bacteria. It possesses a diverse set of virulence factors capable of causing severe and aggressive infections in humans and animals. Antibiotic resistance genes are easily transmitted to humans via contaminated seafood, resulting in a serious antibiotic resistance. The ability of *P. aeruginosa* to form a biofilm maintains its environmental survival and allows its quick adaptation to harsh environments. Therefore, for the benefit of customers and public health globally, the safety and bacteriological quality of commercially processed fish and its products are crucial.

KEYWORDS *P. aeruginosa*, Fish, Antibiotic resistance, Virulence factors, Biofilm

INTRODUCTION

Pseudomonas aeruginosa is a highly pathogenic rod-shaped, Gram-negative bacterium which belongs to family *Pseudomona*daceae, a member of γ -proteobacteria, that is found abundance in water, plants, soil, and animals (Ali *et al.*, 2021). The most frequently isolated bacteria from spoiled seafood are psychrotolerant *Pseudomonas* spp., which produce off flavors and odours as well as slime, which lowers the quality of the product (Benie *et al.*, 2017). The prevalence of *P. aeruginosa* infection, which includes infections of the digestive system, is 17% in underdeveloped nations and 11.5% in Europe, contaminated fish with enterotoxigenic *P. aeruginosa* causes diarrhoea, gastrointestinal disorders, and skin infections, especially in patients who are immunocompromised (Losito *et al.*, 2022; Ziarati *et al.*, 2022).

The pathogenicity of *P. aeruginosa* is associated with the presence of numerous virulence factors, some of which are related to the bacterial cell surface and include lipopolysaccharide, flagella, type IV pili, type III secretion system, exotoxin A, proteases, and alginate, which contribute in the transformation of active proteins as well as the adherence and colonization of bacteria within a host cell (Ali *et al.*, 2023).

P. aeruginosa is known for its resistance to an extensive variety of commonly used antibiotics. As a result, it appears to be more hazardous than other food-spoilage bacteria because it is capable of transmitting multi-drug-resistance (MDR) plasmids to individuals after ingesting infected undercooked fish and fish products containing MDR *P. aeruginosa* (Shahrokhi *et al.*, 2022).

The majority of Gram-positive and Gram-negative bacteria, such as *Escherichia coli*, *Klebsiella pneumoniae*, *Vibrio vulnificus*,

Enterococcus faecalis, Staphylococcus aureus, Streptococcus viridans, Proteus mirabilis, and Pseudomonas aeruginosa, are capable of forming biofilm (MubarakAli et al., 2023). P. aeruginosa is shielded from exposure to atmospheric oxygen and high antibiotic concentrations by biofilms, which are structured communities of bacterial cells that offer increased resistance to environmental stresses (Vetrivel et al., 2021). Since even low concentrations of antibiotics could act as stress signals, biofilm formation is a protective response against the effects of antibiotics. The primary physical obstacle influencing the penetration of antibiotics can be attributed to the biofilm matrix (El Bayomi et al., 2020).

As a result, in this review, the characteristics of *P. aeruginosa* food poisoning public health impacts including contamination of fish and fish products by this important pathogen, virulence factors, antimicrobial resistance profile, and biofilm formation were explored.

Fish and fish products as potential sources of *P. aeruginosa*

The microbiological quality of fish and various types of sea food must be managed due to the rising consumption of these products. Fish is rich in omega 6, omega 3, protein, minerals, and vitamins (Morshdy *et al.*, 2022a). Fish and seafood are more liable to microbial deterioration than other type of meat due to a higher moisture content and a lower pH. Microbial decomposition of fish may cause off flavor, off-odor, slime formation, and discoloration that makes it undesirable for consumption (Morshdy *et al.*, 2019) *Pseudomonadaceae* are aerobic bacteria that are able to develop in the presence of oxygen. However, the total number

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of *Pseudomonas* spp. in a vacuum and CO₂-packed preserved fish is diminished (Lougovois and Kyrana, 2005). *Pseudomonads* are among the most significant specific spoiling organisms (SSOs), as their excessive proliferation accelerates the splitting of nitrogenous materials, which causes product deterioration (Zhang *et al.*, 2021). Numerous studies have shown that Gram-negative bacteria from the *Pseudomonadaceae* family are among the most productive populations of microbes to extracellular proteolytic, lipolytic, and saccharolytic enzymes (Abbamondi and Tommonaro, 2022). The most prevalent *Pseudomonas* species isolated from fish are *P. fluorescens*, *P. lundensis*, *P. fragi*, *P. anguilliseptica*, and *P. putida* (Abd El Tawab *et al.*, 2016). *P. aeruginosa* is the most common *Pseudomonas* species in food products, but it is not widely distributed (Heir *et al.*, 2021).

Pseudomonas aeruginosa Characteristics and Public health impact

P. aeruginosa can survive via water, on various surfaces, and on medical devices with the aid of its powerful adhesion components, including flagella, pili, and biofilm. *P. aeruginosa* is therefore prevalent in both natural and artificial settings, such as lakes, hospitals, and household washbasin drains (Remold *et al.*, 2011). *P. aeruginosa* is an opportunistic pathogen that infects people with a variety of illnesses (Figure 1). *P. aeruginosa* is the second most common pathogen associated with ventilator-related pneumonia, the seventh major causative pathogen of catheter-linked bloodstream infections, and the sixth most frequent organism associated with nosocomial infections (Hidron *et al.*, 2008). In addition, infections at the surgical site, infections of the urinary tract, burn infections of wounds, keratitis, and otitis media (Ito *et al.*, 2021). It is now a significant contributor to resistance to antibiotics and nosocomial illnesses (Shariati *et al.*, 2018).

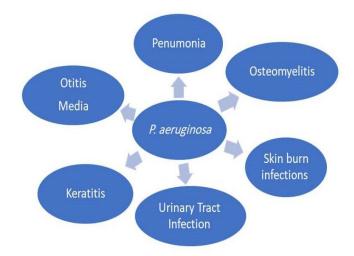


Figure 1. Pseudomonas aeruginosa primary infections

P. aeruginosa is an organism that can quickly acquire antibiotic resistance, adapt to environmental changes, and produce a wide range of virulence factors. The fact that this pathogen is capable of bypassing both innate and acquired immune defenses through adhesion, colonization, and biofilm formation, as well as the production of various virulence factors that cause substantial harm to tissues and this makes it a threat to immunocompromised individuals. Additionally, it contributes to illnesses with high mortality rates in people with cystic fibrosis, infections in newborns, tumors, and serious burns (Nathwani *et al.*, 2014). When insufficient treatment is used to treat *P. aeruginosa* infections, especially when multidrug-resistant (MDR) strains are involved, infections can be fatal (Bail *et al.*, 2022). For the past 30 years, multidrug resistance has posed a threat to both human and animal health (Sarabhai *et al.*, 2013). Additionally, *P. aeruginosa* is one of the most common pathogens in hospital settings and is responsible for more than 50% of healthcare-acquired infections (McKay and Bamford, 2015). Although new antimicrobial medications have been created, *P. aeruginosa* mortality rates remain high, which range from 20 to 60% (Kang *et al.*, 2003). The primary contributory factors of *P. aeruginosa* infections are structural lung diseases, hematological neoplasms, transplantation, burned skin, current antibiotic use, the presence of implants, extended hospitalization, and mechanical ventilation (Reynolds and Kollef, 2021).

Pseudomonas aeruginosa virulence factors

By producing a wide range of virulence factors, P. aeruginosa has the capability to adjust to the unfavorable environment in hosts and enhance the likelihood of illness and infection (Vidaillac and Chotirmall, 2021). First, lipopolysaccharide (LPS) is a crucial surface structural element to safeguard the external host cells toxins. The endotoxicity of the lipid A in LPS facilitates damage to tissues, attachment, and recognition by host receptors (Park et al., 2022). The LPS may be involved in the creation of biofilms and antibiotic resistance (Chambers et al., 2017). The second is that outer membrane proteins (OMPs) support adhesion, exchange of nutrients, and resistance to antibiotics (Sabnis et al., 2021). The third is that the flagellum, pili, and other adhesins are correlated with drug resistance triggered by biofilm formation (Ozer et al., 2021). Fourth, there are six different categories of secretion systems. These include flagella (T6SS-associated), pili (T4SS), and multi-toxin components type 3 secretion system (T3SS), which are used for colonizing the host, adhering to it, swimming, and swarming in response to chemotactic signaling. Finally, exopolysaccharides like alginate, Psl, and Pel may hinder bacterial clearance while aiding in the formation of biofilms (Ozer et al., 2021).

Concerning toxins produced by P. aeruginosa, T3SS is an intricate system that has the potential to seriously impair host defense by injecting cytotoxins such as ExoU, ExoT, ExoS, and ExoY. These toxins have an impact on the intracellular environment, particularly by inhibiting phagocytosis and clearance of bacteria. Exotoxin A (ETA) can prevent the production of host proteins by activating ADP ribosylation (Yang et al., 2022). Additionally, pyocyanin is hazardous to hosts, exacerbates disease, harms host tissue, and negatively impacts organ function (Alatraktchi et al., 2020). In addition, a significant number of lytic enzymes that control additional virulence factors include the elastases LasA and LasB, alkaline protease (AprA) LipC lipases, phospholipase C, and esterase A (Chadha et al., 2022). Furthermore, lung epithelial or tracheal cells can suffer from direct harm from tight junction destruction and lung surfactant degradation caused by rhamnolipids (Zhao et al., 2021). Additionally, siderophores (pyoverdine and pyochelin) function as iron uptake systems to aid bacterial survival in environments with low levels of iron, increasing virulence (Perraud et al., 2022). Finally, in phagocyte environments, reactive oxygen species (ROS) are neutralized by antioxidant enzymes like catalases (KatA, KatB, and KatE), alkyl hydroperoxide reductases, and superoxide dismutases to prevent bacterial clearance (Dar et al., 2021).

Antibiotic resistance in P. aeruginosa

Under increasing stress at aquatic systems, numerous studies show that the usage of antimicrobial agents in fish farming has

been increased as a result. Investigators estimate that by 2030, approximately 13,495 tons of antimicrobials will be applied to aquaculture, corresponding to 5.7% of the total amount used globally (including those utilized in humans and all other animals) (Ben Mhenni et al., 2023). This also attracts consideration to the reality that most antimicrobial categories are employed to treat infections caused by bacteria in both veterinary and human medicine. This accelerates the generation of drug-resistant bacteria, and antimicrobial-resistant bacteria may diminish treatment efficacy, particularly in infections caused by Gram-negative bacteria (Delannoy et al., 2022). Additionally, overusing of antibiotics during therapy hastens the emergence of multidrug-resistant P. aeruginosa strains, rendering antibiotic therapy ineffective against this pathogen (Hirsch and Tam, 2010). As a result, improper application of antibiotics in aquaculture contributes to the increasing prevalence of antimicrobial resistance (AMR), potentially resulting in animal, human, and ecosystem consequences (Schar et al., 2020). According to Hancock and Speert (2000), P. aeruginosa exhibits resistance to a number of antibiotics, including β-lactams aminoglycosides and quinolones. The main defense mechanisms employed by P. aeruginosa against antibiotic outbreak can generally be divided into intrinsic, acquired, and adaptive resistance. Numerous Pseudomonas strains have been found to have a significant degree of intrinsic resistance to the majority of antibiotics (Kačániová et al., 2017). The simultaneously mechanisms of low outer membrane permeability, efflux systems that remove antibiotics from cells, and the synthesis of antibiotic-inactivating enzymes like lactamases are the main contributors to this intrinsic resistance (Breidenstein et al., 2011).

Recent investigations confirmed that various strains of Pseudomonas spp. are able to be resistant to some antimicrobial agents of different classes, particularly lactams such as penicillins, cephalosporins, carbapenems, and monobactams (Elbehiry et al., 2022). According to the World Health Organization, P. aeruginosa is carbapenem-resistant (Karampatakis et al., 2018). In this regard, Kačániová et al. (2019) demonstrated a high percentage of resistant isolates among Pseudomonas spp. identified in fish. Furthermore, all Pseudomonas spp. were meropenem resistant. As reported by Fazeli and Momtaz (2014), Pseudomonas strains displayed the greatest degree of resistance to penicillin (100%), followed by tetracycline (90.19%), streptomycin (64.70%), and erythromycin (43.13%). P. aeruginosa develops a biofilm in the lungs of infected patients, acting as a diffusion barrier to prevent the bacterial cells from being exposed to antibiotics (Drenkard, 2003). The biofilm can also develop multidrug-tolerant persisted cells, which can withstand antibiotic treatment (Mulcahy et al., 2010).

Biofilm formation ability of P. aeruginosa

One of the most important mechanisms for species survival despite unanticipated changes in living conditions, such as temperature and nutrition availability, is the biofilm, which is an extensive collection of bacteria enclosed in a self-generated structure of extracellular polymeric substances (Ahmed *et al.*, 2018; Morshdy *et al.*, 2022b). Due to an increase in rates of mutations, an increase of efflux pumps, a reduction in metabolic activity, and other physical variables, resistance to antibiotics in attached bacteria is approximately 1000 times greater than in planktonic cells (Høiby *et al.*, 2010). *P. aeruginosa* settles a variety of surfaces, such as medical supplies and equipment used in the food industry, and forms biofilms that result in chronic infections because of the organism's increased resistance to antibiotics, different irradiation therapies, environmental factors, disinfecting agents, and

immunity (Stewart and Costerton, 2001).

P. aeruginosa biofilms can typically grow on abiotic surfaces like industrial machinery or medical implants. P. aeruginosa produces infections with the help of numerous virulence factors based on the cell such as lectins, pili, lipopolysaccharide, alginate, and secreted virulence factors (cytotoxin, pyocyanin, proteases, siderophores, hemolysins, exotoxin A, exoenzyme S, exoenzyme U (Strateva et al., 2011). Six separate stages make up the development of the biofilm. Stage 1: Via the assistance of cell appendages such flagella and type IV pili, bacterial cells adhere to a surface (Klausen et al., 2003). Flagellar movement restriction has been linked to twitching motility and the formation of exopolysaccharides required for surface association (Guilbaud et al., 2017), but this adherence is reversible. Stage 2: The transition of bacterial cells from reversible to irreversible adhesion. Stage 3: Progression of connected bacteria into microcolonies, which have a more organized architecture. Stage 4: As a biofilm matures, these microcolonies grow into substantial three-dimensional mushroom-like structures. Stage 5: To release dispersed cells, the matrix cavity in the center of the microcolony is destroyed by cell autolysis (Ma et al., 2009). Stage 6: The biofilm cycle can then repeat after the change from sessile to planktonic growth mode for seeding of uncolonized areas (Rasamiravaka et al., 2015).

L-glucuronic acid and D-mannuronic acid make up the unbranched polymer chain known as alginate. For the biofilm structure to be safeguarded and to remain stable, this polymer is crucial. Alginate helps to keep the matrix's nutrients and water from degrading by preserving its contents (Rasamiravaka *et al.*, 2015). Another significant component of *P. aeruginosa* biofilm is eDNA, which functions as a nutrient base for embedded bacteria and is involved in cell-to-cell communication (Mulcahy *et al.*, 2010). *P. aeruginosa* biofilm structure stability is maintained by many polysaccharides, which comprise pel, alginate, and psl (Ghafoor *et al.*, 2011).

CONCLUSION

Pseudomonas aeruginosa is one of the most prevalent emerging bacteria commonly isolated from fish and fish products. Recovery of multi-drug resistant (MDR) *P. aeruginosa* strains serves as a cautionary tale for the appropriate use of antibiotics. Antimicrobial susceptibility screening must be carried out on a regular basis for the purpose of avoiding the development of antibiotic-resistant strains that could pose a public health concern. The majority of *Pseudomonas* species easily form biofilms and persist as the etiological agent of biofilm-mediated illnesses that result in chronic infectious diseases and recurrent infections. Numerous studies and approaches have been used to control the pathogenesis of biofilm formation and understand the underlying mechanism of biofilm development.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- Abbamondi, G.R.,Tommonaro, G., 2022. Research Progress and Hopeful Strategies of Application of Quorum Sensing in Food, Agriculture and Nanomedicine. Microorganisms 10, 1192.
- Abd El Tawab, A.A., Maarouf, A.A., Ahmed, N.M., 2016. Detection of Virulence factors of *Pseudomonas* species isolated from fresh water fish by PCR. Benha Vet. Med. J. 30, 199-207.
- Ahmed, H.A., El Bayomi, R.M., Hussein, M.A., Khedr, M.H., Remela, E.M.A., El-Ashram, A.M., 2018. Molecular characterization, antibiotic resistance pattern, and biofilm formation of *Vibrio parahaemolyticus* and *V. cholerae* isolated from crustaceans and humans. Int. J.

Food Microbiol. 274, 31-37.

- Alatraktchi, F.A., Svendsen, W.E., Molin, S., 2020. Electrochemical detection of pyocyanin as a biomarker for *Pseudomonas aeruginosa*: A focused review. Sensors 20, 5218.
- Ali, N.G., Ali, T.E.S., Aboyadak, I.M., Elbakry, M.A., 2021. Controlling *Pseudomonas aeruginosa* infection in *Oreochromis niloticus* spawners by cefotaxime sodium. Aquaculture 544, 737107.
- Ali, H., Awad, A., Maarouf, A., Ahmed, W., 2023. Molecular Detection of some Virulence Factors of *Pseudomonas aeruginosa* Isolated from Freshwater Fishes at Qalubiya Governorate, Egypt. Benha Vet. Med. J. 43, 80-84.
- Bail, L., Ito, C.A.S., Arend, L.N.V.S., da Silva Nogueira, K., Tuon, F.F., 2022. Activity of imipenem-relebactam and ceftolozane-tazobactam against carbapenem-resistant *Pseudomonas aeruginosa* and KPC-producing Enterobacterales. Diagn. Microbiol. Infect. Dis. 102, 115568.
- Ben Mhenni, N., Alberghini, G., Giaccone, V., Truant, A., Catellani, P., 2023. Prevalence and Antibiotic Resistance Phenotypes of *Pseudomonas* spp. in Fresh Fish Fillets. Foods 12, 950.
- Benie, C.K.D., Dadié, A., Guessennd, N., N'gbesso-Kouadio, N.A., Kouame, N.Z.D., N'golo, D.C., Dosso, M., 2017. Characterization of virulence potential of *Pseudomonas aeruginosa* isolated from bovine meat, fresh fish, and smoked fish. Eur. J. Microbiol. Immunol. 7, 55-64.
- Breidenstein, E.B., de la Fuente-Núñez, C., Hancock, R.E., 2011. *Pseudomonas aeruginosa*: all roads lead to resistance. Trends in microbiol. 19, 419-426.
- Chadha, J., Harjai, K., Chhibber, S. 2022. Revisiting the virulence hallmarks of *Pseudomonas aeruginosa*: a chronicle through the perspective of quorum sensing. Environ. Microbiol. 24, 2630-2656.
- Chambers, J. R., Cherny, K. E., Sauer, K., 2017. Susceptibility of *Pseudomonas aeruginosa* dispersed cells to antimicrobial agents is dependent on the dispersion cue and class of the antimicrobial agent used. Antimicrob. Agents Chemother. 61, e00846-00817.
- Dar, H.H., Anthonymuthu, T.S., Ponomareva, L.A., Souryavong, A. B., Shurin, G.V., Kapralov, A.O., Tyurin, V.A., Lee, J.S., Mallampalli, R.K., Wenzel, S. E. Bayir, H., Kagan, V.E., 2021. A new thiol-independent mechanism of epithelial host defense against *Pseudomonas aeruginosa*: iNOS/NO• sabotage of theft-ferroptosis. Redox. Biol. 45, 102045.
- Delannoy, S., Hoffer, C., Youf, R., Dauvergne, E., Webb, H. E., Brauge, T., Tran, M., Midelet, G., Granier, S.A., Haenni, M., Fach, P., Brisabois, A., 2022. High Throughput Screening of Antimicrobial Resistance Genes in Gram-Negative Seafood Bacteria. Microorganisms 10, 1225.
- Drenkard, E., 2003. Antimicrobial resistance of *Pseudomonas aeruginosa* biofilms. Microbes infect. 5, 1213-1219.
- El Bayomi, R.M., El Mesalamy, Y., Hafez, A.E., Ahmed, H.A., 2020. Clostridium perfringens in Meat and Meat Products: A minireview on the Incidence, Public Health Significance, and the Effects of Essential Oils. Zag. Vet. J. 48, 340-353.
- Elbehiry, A., Marzouk, E., Aldubaib, M., Moussa, I., Abalkhail, A., Ibrahem, M., Hamada, M., Sindi, W., Alzaben, F., Almuzaini, A.M., Algammal, A.M., Rawway, M. 2022. *Pseudomonas* species prevalence, protein analysis, and antibiotic resistance: An evolving public health challenge. AMB Express 12, 1-14.
- Fazeli, N., Momtaz, H. 2014. Virulence gene profiles of multidrug-resistant *Pseudomonas aeruginosa* isolated from Iranian hospital infections. Iran. Red Crescent Med. J. 16, e15722.
- Ghafoor, A., Hay, I.D., Rehm, B.H., 2011. Role of exopolysaccharides in *Pseudomonas aeruginosa* biofilm formation and architecture. Appl. Environ. Microbiol. 77, 5238-5246.
- Guilbaud, M., Bruzaud, J., Bouffartigues, E., Orange, N., Guillot, A., Aubert-Frambourg, A., Bellon-Fontaine, M.N., 2017. Proteomic response of *Pseudomonas aeruginosa* PAO1 adhering to solid surfaces. Front. Microbiol. 8, 1465.
- Hancock, R. E., Speert, D. P., 2000. Antibiotic resistance in *Pseudomonas aeruginosa*: mechanisms and impact on treatment. Drug resist. updates 3, 247-255]
- Heir, E., Moen, B., Åsli, A. W., Sunde, M., Langsrud, S., 2021. Antibiotic resistance and phylogeny of *Pseudomonas* spp. isolated over three decades from chicken meat in the Norwegian food chain. Microorganisms 9, 207.
- Hidron, A.I., Edwards, J.R., Patel, J., Horan, T.C., Sievert, D.M., Pollock, D.A., Fridkin, S.K., 2008. Antimicrobial-resistant pathogens associated with healthcare-associated infections: annual summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006–2007. Infect. Control. Hosp. Epidemiol. 29, 996-1011.
- Hirsch, E.B., Tam, V.H., 2010. Impact of multidrug-resistant *Pseudomonas* aeruginosa infection on patient outcomes. Expert Rev. Pharma-

coecon. Outcomes Res. 10, 441-451.

- Høiby, N., Bjarnsholt, T., Givskov, M., Molin, S., Ciofu, O., 2010. Antibiotic resistance of bacterial biofilms. Int. J. Antimicrob. Agents 35, 322-332.
- Ito, C.A.S., Bail, L., Arend, L.N.V.S., Nogueira, K.D.S., Tuon, F.F., 2021. The activity of ceftazidime/avibactam against carbapenem-resistant *Pseudomonas aeruginosa*. Infect. Dis. 53, 386-389.
- Kačániová, M., Klūga, A., Kántor, A., Medo, J., Žiarovská, J., Puchalski, C., Terentjeva, M., 2019. Comparison of MALDI-TOF MS Biotyper and 16S rDNA sequencing for the identification of *Pseudomonas* species isolated from fish. Microb. Pathog. 132, 313-318.
- Kačániová, M., Terentjeva, M., Vukovic, N., Puchalski, C., Roychoudhury, S., Kunová, S., Klūga, A., Tokár, M., Kluz, M., Ivanišová, E., 2017. The antioxidant and antimicrobial activity of essential oils against *Pseudomonas* spp. isolated from fish. Saudi Pharm. J. 8, 1108-1116.
- Kang, C.-I., Kim, S.-H., Kim, H.-B., Park, S.-W., Choe, Y.-J., Oh, M.-d., Kim, E.-C, Choe, K.-W., 2003. *Pseudomonas aeruginosa* bacteremia: risk factors for mortality and influence of delayed receipt of effective antimicrobial therapy on clinical outcome. Clin. Infect. Dis. 6, 745-751.
- Karampatakis, T., Antachopoulos, C., Tsakris, A., Roilides, E., 2018. Molecular epidemiology of carbapenem-resistant *Pseudomonas aeruginosa* in an endemic area: comparison with global data. Eur. J. Clin. Microbiol. Infect. Dis. 37, 1211-1220.
- Klausen, M., Aaes-Jørgensen, A., Molin, S., Tolker-Nielsen, T., 2003. Involvement of bacterial migration in the development of complex multicellular structures in *Pseudomonas aeruginosa* biofilms. Mol. Microbiol. 50, 61-68.
- Losito, A. R., Raffaelli, F., Del Giacomo, P., Tumbarello, M., 2022. New drugs for the treatment of *Pseudomonas aeruginosa* infections with limited treatment options: A narrative review. Antibiotics 11, 579.
- Lougovois, V., Kyrana, V., 2005. Freshness quality and spoilage of chillstored fish. Food Policy, Control Res. 1, 35-86.
- Ma, L., Conover, M., Lu, H., Parsek, M.R., Bayles, K., Wozniak, D.J., 2009. Assembly and development of the *Pseudomonas aeruginosa* biofilm matrix. PLoS pathog. 5 , e1000354]
- McKay, R., Bamford, C., 2015. Community-versus healthcare-acquired bloodstream infections at groote schuur hospital, cape town, South Africa. S. Afr. Med. J. 105, 363-369.
- Morshdy, A.E.M., Darwish, W.S., Daoud, J.R.M., Sebak, M.A.M., 2019. Estimation of metal residues in *Oreochromis niloticus* and Mugil cephalus intended for human consumption in Egypt: a health risk assessment study with some reduction trials. J. Consum. Prot. Food Saf. 14, 81-91.
- Morshdy, A.E.M., Abdelhameed, N.S.A., El Bayomi, R.M., Abdallah, K., 2022a. Prevalence of Antibiotic Resistant *Aeromonas* and Molecular Identification of *Aeromonas hydrophila* Isolated from Some Marketed Fish in Egypt. J. Adv. Vet. Anim. Res. 12, 717-721.
- Morshdy, A.E.M., El-Tahlawy, A.S., Qari, S.H., Qumsani, A.T., Bay, D.H., Sami, R., Qumsani, A.T., Bay, D.H., Sami, R., Althubaiti, E.H., Mansour, A.M.A., Aljahani, A.H., Hafez, A.E., Mahmoud, A.F.A., El Bayomi, R.M., Hussein, M.A., 2022b. Anti-Biofilms' Activity of Garlic and Thyme Essential Oils against *Salmonella* Typhimurium. Molecules 27, 2182.
- MubarakAli, D., Arunachalam, K., Lakshmanan, M., Badar, B., Kim, J.-W., Lee, S.-Y., 2023. Unveiling the Anti-Biofilm Property of Hydroxyapatite on *Pseudomonas aeruginosa*: Synthesis and Strategy. Pharmaceutics 15, 463.
- Mulcahy, H., Charron-Mazenod, L., Lewenza, S., 2010. Pseudomonas aeruginosa produces an extracellular deoxyribonuclease that is required for utilization of DNA as a nutrient source. Environ. Microbiol. 12, 1621-1629.
- Nathwani, D., Raman, G., Sulham, K., Gavaghan, M., Menon, V., 2014. Clinical and economic consequences of hospital-acquired resistant and multidrug-resistant *Pseudomonas aeruginosa* infections: a systematic review and meta-analysis. Antimicrob. Resist. Infect. Control 3, 1-16.
- Ozer, E., Yaniv, K., Chetrit, E., Boyarski, A., Meijler, M.M., Berkovich, R., Kushmaro, A., Alfonta, L., 2021. An inside look at a biofilm: *Pseudomonas aeruginosa* flagella biotracking. Sci. Adv. 7, eabg8581.
- Park, W. S., Lee, J., Na, G., Park, S., Seo, S.-K., Choi, J. S., Jung, W.-K., Choi, I.-W., 2022. Benzyl Isothiocyanate Attenuates Inflammasome Activation in *Pseudomonas aeruginosa* LPS-Stimulated THP-1 Cells and Exerts Regulation through the MAPKs/NF-κB Pathway. Int. J. Mol. Sci. 23, 1228.
- Perraud, Q., Kuhn, L., Fritsch, S., Graulier, G., Gasser, V., Normant, V., Hammann, P., Schalk, I.J., 2022. Opportunistic use of catecholamine neurotransmitters as siderophores to access iron by *Pseudomo-*

nas aeruginosa. Environ. Microbiol. 24, 878-893.

- Rasamiravaka, T., Labtani, Q., Duez, P., El Jaziri, M., 2015. The formation of biofilms by *Pseudomonas aeruginosa*: a review of the natural and synthetic compounds interfering with control mechanisms. BioMed. Res. Int. 2015, 1-18.
- Remold, S.K., Brown, C.K., Farris, J.E., Hundley, T.C., Perpich, J.A., Purdy, M.E., 2011. Differential habitat use and niche partitioning by *Pseudomonas* species in human homes. Microb. Ecol. 62, 505-517.
- Reynolds, D., Kollef, M., 2021. The epidemiology and pathogenesis and treatment of *Pseudomonas aeruginosa* infections: an update. Drugs 81, 2117-2131.
- Sabnis, A., Hagart, K.L., Klöckner, A., Becce, M., Evans, L.E., Furniss, R.C.D., Mavridou, D., Murphy, R., Davies, J.C., 2021. Colistin kills bacteria by targeting lipopolysaccharide in the cytoplasmic membrane. eLife 10, e65836.
- Sarabhai, S., Sharma, P., Capalash, N., 2013. Ellagic acid derivatives from Terminalia chebula Retz. downregulate the expression of quorum sensing genes to attenuate *Pseudomonas aeruginosa* PAO1 virulence. PLoS one 8, e53441.
- Schar, D., Klein, E.Y., Laxminarayan, R., Gilbert, M., Van Boeckel, T.P., 2020. Global trends in antimicrobial use in aquaculture. Sci. Rep. 10, 21878.
- Shahrokhi, G. R., Rahimi, E., Shakerian, A., 2022. The prevalence rate, pattern of antibiotic resistance, and frequency of virulence factors of *Pseudomonas aeruginosa* strains isolated from fish in Iran. J. Food Qual. 2022, 1-8.
- Shariati, A., Azimi, T., Ardebili, A., Chirani, A., Bahramian, A., Pormohammad, A., Sadredinamin, M., Erfanimanesh, S., Bostanghadiri, N., Shams, S. Hashemi, A., 2018. Insertional inactivation of oprD in

carbapenem-resistant *Pseudomonas aeruginosa* strains isolated from burn patients in Tehran, Iran. New Microb. New Infect. 21, 75-80.

- Stewart, P.S., Costerton, J.W., 2001. Antibiotic resistance of bacteria in biofilms. The lancet 358, 135-138.
- Strateva, T., Mitov, I., 2011. Contribution of an arsenal of virulence factors to pathogenesis of *Pseudomonas aeruginosa* infections. Ann. Microbiol. 61, 717-732.
- Vetrivel, A., Ramasamy, M., Vetrivel, P., Natchimuthu, S., Arunachalam, S., Kim, G.-S., Murugesan, R., 2021. *Pseudomonas aeruginosa* biofilm formation and its control. Biologics 1, 312-336.
- Vidaillac, C., Chotirmall, S.H., 2021. Pseudomonas aeruginosa in bronchiectasis: infection, inflammation, and therapies. Expert Rev. Respir. Med. 15, 649-662.
- Yang, J.J., Tsuei, K.-S.C., Shen, E.P., 2022. The role of Type III secretion system in the pathogenesis of *Pseudomonas aeruginosa* microbial keratitis. Tzu-Chi Med. J. 34, 8-14.
- Zhang, Z., Wu, R., Gui, M., Jiang, Z., Li, P., 2021. Identification of the specific spoilage organism in farmed sturgeon (*Acipenser baerii*) fillets and its associated quality and flavour change during ice storage. Foods 10, 2021.
- Zhao, F., Wang, Q., Zhang, Y., Lei, L., 2021. Anaerobic biosynthesis of rhamnolipids by *Pseudomonas aeruginosa*: performance, mechanism and its application potential for enhanced oil recovery. Microb. Cell Factories 20, 1-12.
- Ziarati, M., Zorriehzahra, M.J., Hassantabar, F., Mehrabi, Z., Dhawan, M., Sharun, K., Emran, T.B., Dhama, K., Chaicumpa, W., Shamsi, S., 2022. Zoonotic diseases of fish and their prevention and control. Vet. Q. 42, 95-118.