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Prevalence of Mould and Aflatoxin in Raw and Heat-Treated Meat Products

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INTRODUCTION

Meat products are a rich source of concentrated nutrients since they include protein with a high digestibility score as well as vitamins, minerals, vital amino acids, and fatty acids that are believed to be important for healthy human growth in both young and adult populations. Moreover, provide clients with a source for fast, inexpensive, and nutritious meals, which they highly value for its flavor, affordability, and simplicity of preparation (Hussein et al., 2018). The Inadequate sanitary and hygienic conditions during handling, processing, and storage were the cause of meat product mold contamination. It is possible to enhance the amount of mold contamination in such items by adding flavoring agents of poor quality. Spices and other flavorings added to the meat product mix can significantly enhance the meat's mold contamination (Habashy et al., 2019). Mycotoxins are a wide and potent class of poisonous chemicals with adverse effects on both humans and animals (Bennett and Klich, 2003). They contaminate a wide range of foods and are secondary fungal compounds generated during mould development. Grains, rice, beans, coffee, wine, fruits, nuts, spices, eggs, and animal products are among the foods considered to be the riskiest. The issue is that, despite research efforts and mitigation measures, their occurrence is not entirely prevented. Carcinogenicity, teratogenicity, immunological toxicity, neurotoxicity, hepatotoxicity, nephrotoxicity, reproductive and developmental toxicity, indigestion, and other

negative consequences have been seen in both people and animals (Pleadin et al., 2019). Food safety and hygiene can still be compromised by mycotoxins, viruses, and bacteria (De Ruyck et al., 2015). Although the FAO reported that cereals were 25% mycotoxin-contaminated in 1999, more current statistics reveal that contamination to be substantially greater (about 60-80%) (Eskola et al., 2019). Studies have shown that there are various variables, including the kind of mycotoxin and the analytical or reporting methods employed, that affect the worldwide mycotoxin prevalence in crops. In addition, the vast 2008-2017 research on various cereals and their derivatives conducted in almost a hundred nations demonstrated a high correlation between mycotoxin incidence and meteorological circumstances. Most samples (88% were positive for at least one mycotoxin) and their co-occurrence (64% of samples were positive for at least two mycotoxins) were found to contain mycotoxins (Gruber-Dorninger et al., 2019). According to evidence, there are three ways that meat products can become contaminated: (i) through contaminated raw materials like spices and other ingredients; (ii) through mycotoxin-producing molds found on the surface of dry-cured meat products; and (iii) through the spillover effect from farm animals exposed to contaminated feed (Asefa et al., 2011; Bertuzzi et al., 2013; Pleadin et al., 2013). Mycotoxins frequently linger in raw materials and finished goods and build up in the body of humans, leading to serious health problems that are brought on by eating contaminated food (Richard, 2007; Duarte et al., 2010). The current

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Abstract

From several stores and butchers in Mansoura city, Dakahlia Governorate, Egypt, 120 samples of sausage, beef burger, minced meat, luncheon, hot dog and canned meat were collected (20 each). The samples examined for detection of total mould count and identification if mould into genera and species in addition to quantification of aflatoxin B1, B2, G1 and G2. Mould detected in 100% of all examined raw meat products meanwhile, detected in 25%, 30% and 15% of examined luncheon, hotdog, and canned meat, respectively. Heat treated meat products significantly (P<0.05) contained lower mould count than raw meat products. Eight mould genera detected in all examined meat products with varying percentages in descending order *Aspergillus > Penicillium > Cladosporium > Sporotricum > Alternaria > Mucor > Fusarium > Curvularia*. The mean values of aflatoxin B1 were 0.78±0.21, 1.1±0.55, 1.54±0.40, 0.052±0.032, 2.21±0.87 and 1.88±0.41 μ g/kg in sausage, beef burger, minced meat products in level of aflatoxin B1(P<0.05). Aflatoxin B2 and G2 not detected in all examined samples. The aflatoxin G1 detected in two samples of beef burger with a mean value 1.15±0.065 μ g/kg and in one sample of canned meat 0.62 μ g/kg. A food safety management system as hazard analysis and critical control points should be adopted by meat produces in order to protect human health.

KEYWORDS Aflatoxin, Aspergillus, Penicillium, Meat products study was designed to evaluate the level of mould contamination in addition to determination of aflatoxin residues in raw and heat-treated meat products.

MATERIALS AND METHODS

Collection and preparation of samples

One hundred twenty samples of sausage, beef burger, minced meat, luncheon, hot dog and canned meat (20 each) were chosen at random from several stores and butchers in Mansoura city, Dakahlia Governorate, Egypt. The samples collected from April through August 2022. In order to evaluate samples mycologically without undue delay as well as detection of aflatoxin, samples were immediately moved to the laboratory of Food Control Department, Faculty of Veterinary Medicine, Zagazig University, under completely aseptic conditions after being stored in sterile polyethylene bags and kept in an ice box. A total of 25 g from each sample were aseptically homogenized at 2500 rpm for two minutes in 225 ml of 0.1% sterile peptone water using a sterile homogenizer. As a result, homogenate indicates a dilution of 10-1, and decimal dilutions were subsequently performed (APHA, 2001).

Estimation and Identification of mould count

Total mold count was calculated by growing duplicate plates on several types of malt extract agar media (MEA) (Oxoid) and incubating them at 25°C for one to seven days. Every day throughout the incubation period, the plates were checked for the development of the star-shaped mold (APHA, 2001). Under aseptic circumstances, mold colonies were collected, then subcultured on MEA slopes and stored for further analysis. The identification of the mold colonies was done by meticulous observation and measurements of the colonies' macroscopic and microscopic features, which were then documented on data sheets (Pitt and Hocking, 2009). The uniformity of the surface growth, the pattern of folding (rugae), the clarity of the colony edge, and the presence of pigment on the colony's surface, its reversal, or dilating into the surrounding media are all things that may be observed during a macroscopical study. A magnifying hand lens was used to view the colony's front and rear sides. microscopical analysis A portion of the colony was quickly stained with a few drops of lactophenol cotton blue stain using mycological needles, then covered with a clean cover slide. The micromorphological characteristics of the head, vesicle, sterigmata, conidiophore, and conidia were evaluated on each slide using low power and high-power magnifying lenses.

Determination of aflatoxin residues

The standards of aflatoxin (B1, B2, G1 and G2) were obtained from Biocomma limited, China. The column capacity and recovery as shown in Table 1. The sample was divided into 100 grams, homogenized, and thoroughly mixed with 10 ml of 20% citric acid made by adding 200 ml of dichloromethane. The mixture was then shaken on a timer for 30 minutes. The mixture was separated, and the components that had been filtered were then vacuum-evaporated. Finally, the extracted material was given hexane. Solid-phase extraction (SPE) columns, Bond Elut C18 (500 mg, 3 ml, or 6 ml; Varian, Les Ulis, France), were used for cleanup. To remove lipids, the extracted materials were added to the gel on top of the column and eluted with hexane. A 1:3:6 ratio of hexane, ether, and acetonitrile was used to eliminate further contaminants (Herzallah, 2009). To acquire or recover aflatoxins, the column was eluted using an elution solution that was a combination of dichloromethane and acetone. Nitrogen evaporators (Turbo Vap® LV, Caliper) were used to evaporate the organic solvent until it was completely dry. Using a gradient approach with a flow rate of 1 ml/min at a temperature of 30°C, 20 microliters of the solution was injected into HPLC along with an isocratic mobile phase made up of deionized water, acetonitrile, and methanol (60:20:20 v/v/v). A reversed-phase column (Extend-C18, Zorbax column, 4.6 mm 250 mm, 5 m, Agilent Co.) was used for the separation. A fluorescence detector with wave lengths of 360 nm excitation and 440 nm emission was used to do the detection. The area under the curves, which was automatically extrapolated using ChemStation software, was used to extract and compute the quantity of residues in the samples.

Statistical analysis

The one-way ANOVA significant at P < 0.05 and Pearson correlation tests were used to evaluate the data.

RESULTS

According to Table 2, mould detected in 100% of all examined

Fraction	Addition level —	Testing results (ng)					\mathbf{D}
		Rep.2	Rep.2	Rep.3	Avg.	RSD (%)	– Recovery (%)
B1	77.256 ng	76.2	79.5	75.4	77	2.8	99.7
B2	16.488 ng	143	13.8	14	14	1.8	84.9
G1	82.224 ng	75.9	77	79.1	77.3	2.1	94
G2	24.408 ng	20.3	19.3	20	19.9	2.6	81.5

Table 1. Column capacity, recovery, and rate of standard deviation of aflatoxin B1, B2, G1 and G2

Table 2. Prevalence and count of mould log10CFU/g in examined raw and heat-treated meat products (N= 20 for each).

	Raw meat products			Heat treated meat products			
	Sausage	Beef burger	Minced meat	Luncheon	Hot dog	Canned meat	
Prevalence	(20/20) 65%	(20/20) 85%	(20/20) 45%	(5/20) 25%	(6/20) 30%	(3/20) 15%	
Minimum	2.36	2.21	1.24	2.1	1.24	1.25	
Maximum	4.21	5.2	3.33	2.44	2.54	2.65	
Mean±SD	3.32±0.71ª	3.50±0.91ª	2.43 ± 0.58^{b}	1.91±0.62°	1.95±0.52°	1.42±0.65°	

 $(^{a, b, c})$ different superscript letters in the same row indicate significant differences (p < 0.05).

raw meat products meanwhile, detected in 25%, 30% and 15% of examined luncheon, hotdog, and canned meat, respectively. Heat treated meat products significantly (P<0.05) contained lower mould count than raw meat products. Eight mould genera detected in all examined meat products with varying percentages in descending order Aspergillus > Penicillium > Cladosporium > Sporotricum > Alternaria > Mucor > Fusarium > Curvularia (Table 3). The recorded data in Table 4 declared that A.niger and A. flavus detected in 40% and 20%, 35% and 30%, 25% and 10% in sausage, beef burger, respectively. The A. fumigatus detected in 10%, 10%, 5% and 5% of examined sausage, beef burger, minced meat and luncheon, respectively. The A. ochraceus detected in 10% of examined sausage and beef burger while detected in 5% of examined luncheon, hot dog and canned meat. The mean values of aflatoxin B1 were 0.78±0.21, 1.1±0. 55, 1.54±0.40, 0.052±0.032, 2.21±0.87 and 1.88±0.41 µg/kg in sausage, beef burger, minced meat, luncheon, hot dog and canned meat, respectively. Minced meat significantly lower than other examined meat products in level of aflatoxin B1(P< 0.05). Aflatoxin B2 and G2 not detected in all examined samples. The aflatoxin G1 detected in two samples of beef burger with a mean value 1.15±0.065 µg/kg and in one sample of canned meat 0.62 μ g/kg (Table 5).

on the globe. Not only producing economic losses but also creating risks for both humans and animals. One of the indicative indicators of a product's hygienic status, which describes the environment, the state, and the circumstances surrounding the production process, is the presence of mold in the meat. In the current study, as recorded in Table 2 the total mold count of sausage (3.32±0.71 log₁₀CFU/g) comparable mould counts 3.08 log-₁₀CFU/g (El Bayomi *et al.*, 2021), 3.04±2.15 log₁₀CFU/g (Abuzaid *et* al., 2020), 3.12±1.23 log₁₀CFU/g (Hamad et al., 2021). Meanwhile, higher mould count in sausage 4.6±4.1 log₁₀CFU/g (Abdel Gawaad and El Leboudi, 2005). The mean value of mould was 3.50±0.91 log₁₀CFU/g in examined beef burger samples which slightly higher than 2.85 log₁₀ CFU/g (El Bayomi et al., 2021), 2.87±1.79 log₁₀ CFU/g (Hamad et al., 2021) and 2.84 log₁₀ CFU/g (Maktabi et al., 2016). The mould count in minced meat was 2.43±0.58-log₁₀ CFU/g which come in consistent with 2.24 log₁₀ CFU/g (El Bayomi et al., 2021), 2.24±1.72 log₁₀ CFU/g (Algammal et al., 2021) and 2.4±2.0 log₁₀ CFU/g (Saad et al., 2015). Meanwhile, higher count 3.2±2.8 log₁₀ CFU/g (Hassan et al., 2014). The mould count in luncheon and hot dog samples slightly lower than 2.27 log₁₀ CFU/g (Ebraheem and Ghadam, 2015) and 2.74 \pm 1.3 log₁₀ CFU/g (Hamad et al., 2021). The mould count in canned meat was lower than 3.28 log₁₀ CFU/g in canned meat from Kingdom Saudi Arabia (Nasser, 2015).

The mould species isolated fromsausage, beef burger, minced meat, luncheon, hot dog and canned meat were nearly similar with those obtained in national studies (Abdel Gawaad and El Leboudi, 2005; Saad *et al.*, 2015; Habashy *et al.*, 2019; El Bayomi *et al.*, 2021) and international studies (Nasser, 2015; Maktabi *et al.*,

DISCUSSION

Mold contamination of food is a severe problem everywhere

Table 3. The number and proportion	of identified mould genera in e	examined raw and heat-treated	meat products ($N=20$ for each).

	Raw meat products			Heat treated meat products			
	Sausage	Beef burger	Minced meat	Luncheon	Hot dog	Canned meat	
Aspergillus	17 (85%)	18 (90%)	11 (55%)	3(15%)	2 (10%)	1 (5%)	
Penicillium	11 (55%)	9 (45%)	6 (30%)	2 (10%)	1 (5%)	-	
Cladosporium	8 (40%)	9 (45%)	3 (15%)	-	2 (10%)	2 (10%)	
Sporotricum	6 (30%)	7 (35%)	2 (10%)	-	-	1 (5%)	
Alternaria	3 (15%)	4 (20%)	-	1 (5%)	2 (10%)	1 (15%)	
Mucor	2 (10%)	3 (15%)	1 (5%)	1 (5%)	-	-	
Fusarium	-	2 (10%)	-	1 (5%)	-	-	
Curvularia	2 (10%)	1 (5%)	1 (5%)	-	1 (5%)	1 (5%)	

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Table 4 The number and proportion of identified Asperditus species	s in examined raw and near-ireated mear products $(n=70)$
Table 4. The number and proportion of identified Aspergillus species	in examined fait and near treated mear products (if 20).

G	Raw meat products			Heat treated meat products			
Species	Sausage	Beef burger	Minced meat	Luncheon	Hot dog	Canned meat	
A. niger	8 (40%)	7 (35%)	5 (25%)	1 (5%)	-	-	
4. flavus	4 (20%)	6 (30%)	2 (10%)	-	1(5%)	-	
4. fumigatus	2 (10%)	2 (10%)	1(5%)	1(5%)	-	-	
4. ochraceus	2 (10%)	2 (10%)	-	1(5%)	1(5%)	1(5%)	
4. terreus	1 (5%)	-	-	-	-	-	
A.parasiticus	-	1 (5%)	3(15%)	-	-	-	

Table 5. Aflatoxin residues (µg/kg) in examined raw and heat-treated meat products (n=5).

	Raw meat products			Heat treated meat products			
	Sausage	Beef burger	Minced meat	Luncheon	Hot dog	Canned meat	
B1	1.14 -3.14 1.88±0.41 ^{ab}	1.25- 4.12 2.21±0.87ª	0.010-0.016 0.052±0.032°	$\begin{array}{c} 0.9\text{-}2.18 \\ 1.54{\pm}0.40^{\mathrm{ab}} \end{array}$	0.98-2.14 1.1±0. 55 ^b	0.38-0.85 0.78±0.21 ^b	
B2	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>	
G1	0.95	1.11- 1.19 1.15±0.065	<lod< td=""><td><lod< td=""><td><lod< td=""><td>0.62</td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td>0.62</td></lod<></td></lod<>	<lod< td=""><td>0.62</td></lod<>	0.62	
G2	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""><td><lod< td=""></lod<></td></lod<></td></lod<>	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>	

 $^{(a,b,c)}$ different superscript letters in the same row indicate significant differences (p < 0.05). <LOD: below the limit od detection.

2016; Zadravec et al., 2020). The mould count Significantly, lower (P< 0.05) in heat-treated meat products than raw meat products come in consistency with fact that most molds are heat-sensitive and affected with heat treatments between 60-71°C (Breidt and Costilow, 2004). Meanwhile presence of mould in heat-treated product with a few percentages and counts may attributed to post processing contamination or and inadequately heat-treated as recommended. The sausage and beef burgers had the highest level of contamination because of the ingredients in both products, which are minced meat blended with different spices, binders, and extenders. These ingredients are typically imported from developing nations with tropical and subtropical climates, where hot temperatures, copious amounts of rain, and humidity frequently encourage fungal growth and facilitate the occurrence of mycotoxin (Pickova et al., 2020). There are multifactorial causes affect the level of meat product contamination during processing as grinding, mincing, cooking, chilling, and packing. Mold may readily get into the meat products through all of these procedures. Therefore, the presence of mold in meat products indicates that these items were prepared in unhygienic processing.

Aflatoxin accumulation in the liver after being consumed in contaminated food, even in little levels, and has a carcinogenic impact. Given that ingesting 28 mg of AFB1 over a lifetime can cause cancer, even extremely low concentration levels (1 ppb) would pose a serious risk to the public's health (Garner, 1992). The most toxic mycotoxin for both people and animals, aflatoxin B1 (AFB1), has been designated as a Group 1 human carcinogen (IARC, 2002). The studied samples varied significantly from one another (P< 0.05). Furthermore, the descending manner for AflatoxinB1 was arranged as follow beef burger > sausage > luncheon > hot dog > canned meat > minced meat. The aflatoxin B1 seems to be correlated with number of spices and additives that had previously been contaminated with aflatoxins may be the reason why the lowest amount of aflatoxin B1 was found in minced meat. Previous national studies declared the level of aflatoxin B1 as 10.4±5.1, 2.3±0.4 ppb in luncheon, basterma (Ismail *et al.*, 213) and 7.23 ± 0.8, 5.63 ± 0.95, 4.88 ± 0.11, 2.03 ± 0.3 ppb luncheon, hot dog, corned beef, and minced meat, respectively (Algahtani et al., 2020). Furthermore, aflatoxin B1 detected globally at level 7-8 ppb in Italy from sausage (lacumin et al., 2009), 1.9-6.3 ppb from dry-cured Iberian ham in Spain (Rodríguez et al., 2012), and up to 1.92 in traditional meat products from Croatia (Zadravec et al., 2020). All examined samples were below the maximum permissible limit for aflatoxin B1 (5 ppb) as maximum level established as regulation for European countries (EC, 2006). Regarding the aflatoxin G1level which come lower than 4.37 ± 0.63 , 5.63 ± 0.2 , 6.9 ± 0.63 and 5.53 ± 0.1 ppb in luncheon, hot dog, corned beef, and minced meat, respectively (Algahtani et al., 2020). Aflatoxin B2 and G2 were not detected in all examined samples on contrary detected as 5.20±0.69 and 3.35±0.49 in sausage, 5.57±0.72 and 3.84±0.58 ppb in luncheon (Shaltout et al., 2014).

CONCLUSION

Meat products in this investigation were contaminated to different mould and aflatoxin B1 which indeed below the permissible limit but long-term consumption may lead to public health hazard. In order to maintain hygienic conditions during processing, preparation, and handling, a concentrated effort should be made. This should be avoided by using hygienic procedures while slaughtering of animals and preparation of meat, selection of species and additive, proper packaging and cooling of raw meat products, proper heat treatment of heat-treated products and training of meat factory workers.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

- Abdel Gawaad, M.I., El Leboudi S.H.E., 2005. Some chemical and mycological examinations of meat and fish products. Vet. Med. J. Giza 55, 941-948.
- Abuzaid, K., Shaltout, F., Salem, R., El-Diasty, E., 2020. Microbial aspect of some processed meat products with special reference to aflatoxins. BVMJ. 39, 24-28.
- Algahtani, F.D., Morshdy, A.E., Hussein, M.A., Abouelkheir, E.S., Adeboye, A., Valentine, A., Elabbasy, M.T., 2020. Biogenic amines and aflatoxins in some imported meat products: Incidence, occurrence, and public health impacts. Journal of Food Quality 2020, 1-7.
- Algammal, A.M., Elsayed, M.E., Hashem, H.R., Ramadan, H., Sheraba, N.S., El-Diasty, E.M., Hetta, H.F., 2021. Molecular and HPLC-based approaches for detection of aflatoxin B 1 and ochratoxin A released from toxigenic *Aspergillus* species in processed meat. BMC Microbiol. 21, 1-10.
- APHA (American Public Health Association), 2001. Compendium of methods for the microbiological examination of foods. 4th Ed. Eds. Downes, F.P. and K. Ito. Sheridan Books Inc., Washington D.C., USA.
- Asefa, D.T., Kure, C.F., Gjerde, R.O., Langsrud, S., Omer, M.K., Nesbakken, T., Skaar, I. A., 2011. HACCP plan for mycotoxigenic hazards associated with dry-cured meat production processes. Food Control 22, 831–837.
- Bennett, J.W., Klich, M., 2003. Mycotoxins. Clin. Microbiol. Rev. 16, 497– 516.
- Bertuzzi, T., Gualla, A., Morlacchini, M., Pietri, A., 2013. Direct and indirect contamination with ochratoxin A of ripened pork products. Food Control 34, 79–83.
- Breidt J. F., Costilow, R.N., 2004. Processing and safety. Foods: Principles of Handling and Preservation 5, 5-16.
- De Ruyck, K., De Boevre, M., Huybrechts, I., De Saeger, S., 2015. Dietary mycotoxins, co-exposure, and carcinogenesis in humans: short review. Mutat. Res. Rev. Mutat. Res. 766, 32–41.
- Duarte, S.C., Pena, A., Lino, C.M., 2010. Ochratoxin A in Portugal: A review to assess human exposure. Toxins 2, 1225–1249.
- Ebraheem, L.M, Ghadam, M., 2015. Evaluation of mycological status and detection of its toxins in basterma and luncheon in assuit city. Assiut Vet. Med. J. 58, 1-10.
- El Bayomi, R.M., Hebishy, R.M., Darwish, W.S., El-Atabany, A.I.M., Mahmoud, A. F.A., 2021. Mould contamination of some meat products with reference to decontamination trials of *Aspergillus flavus* using essential oils. Slov. Vet. Res. 58, 363-372.
- Eskola, M., Kos, G., Elliott, C.T., Hajšlová, J., Mayar, S., Krska, R., 2019. Worldwide contamination of food crops with mycotoxins: Validity of the widely cited 'FAO estimate' of 25%. Crit. Rev. Food Sci. Nutr. 60, 2773–2789.
- Garner, R.1992. Aflatoxin-a cancer problem that refuses to go away. In: Food Safety and Quality Assurance Applications of Immunoassay Systems, M. R. A. Morgan, C. J. Smith, and P. A. Williams, Eds., Vol. 93. Elsevier Science Publishers Ltd, Berlin, Germany. p. 102
- Gruber-Dorninger, C., Jenkins, T., Schatzmayr, G., 2019. Global Mycotoxin Occurrence in Feed: A Ten-Year Survey. Toxins 11, 375.
- Habashy, A., Darwish, W., Hussein M., El-Dien, W., 2019. Prevalence of different mould genera in meat and meat products with some reduction trials using essential oils. Adv. Anim. Vet. Sci. 7, 79-85.
- Hamad, G.M., Mohdaly, A.A., El-Nogoumy, B.A., Ramadan, M.F., Hassan, S.A., Zeitoun, A.M., 2021. Detoxification of aflatoxin B1 and ochratoxin A using salvia farinacea and Azadirachta indica water extract and application in meat products. Appl. Biochem. Biotechnol. 193, 3098-3120.
- Hassan, M., Abd El Aziz, A., Tartor, Y., Farouk, S., 2014. Aflatoxin producing moulds and aflatoxin residues in meat and meat products in Egypt. ZVMJ. 42, 43-55.
- Herzallah, S.M., 2009. Determination of aflatoxins in eggs, milk, meat and meat products using HPLC fluorescent and UV detectors. Food Chem. 114, 1141-1146.
- Hussein, M.A., Eldaly, E.A., Seadawy, H.G., El-Nagar, E.F., 2018. Virulence and antimicrobial resistance genes of *Escherichia coli* in ready to eat sandwiches in Sharkia governorate . Slov. Vet. Re. 383-392.
- Iacumin, L., Chiesa, L., Boscolo, D., Manzano, M., Cantoni, C., Orlić, S., Comi, G., 2009. Moulds and ochratoxin A on surfaces of artisanal and industrial dry sausages. Food Microbiol. 26, 65–70.
- IARC, 2002. Some Traditional Herbal Medicines, Some Mycotoxins, Naphthalene and Styrene, World Health Organization, Geneva, Switzerland.
- Ismail, S.A., Shehata, A.A., El-Diasty, E.M., 2013. Microbiological quality of some meat products in local markets with special reference to mycotoxins. Global Veterinaria 10, 577-584.

- Maktabi, S., Fazlara, A., Ghorbanpoor, M., Talayol, G., Siavashi, M., 2016. Measurement and assessment of aflatoxin B1 and its producing molds in Iranian sausages and burgers. J. kermanshah Univ. Med. Sci. 20, 74-78.
- Nasser L.A., 2015. Molecular identification of isolated fungi, microbial and heavy metal contamination of canned meat products sold in Riyadh, Saudi Arabia. Saudi J. Biol. Sci. 22, 513–520.
- Pickova, D., Ostry, V., Malir, J., Toman, J., Malir, F., 2020. Review on Mycotoxins and Microfungi in Spices in the Light of the Last Five Years. Toxins 12, 789-795.
- Pitt, J.I., Hocking, A.D., 2009. Fungi and Food Spoilage, 3rd Ed. Published by Blackie academic and professional academic press New York, London. https://doi.org/10.1007/978-0-387-92207-2.
- Pleadin, J., Perši, N., Kovačević, D., Vahčić, N., Scortichini, G., Milone, S., 2013. Ochratoxin A in traditional dry-cured meat products produced from subchronic-exposed pigs. Food Addit. Contam. Part A. 30, 1827–1836.
- Pleadin, J.; Frece, J.; Markov, K., 2019. Mycotoxins in Food and Feed. In Advances in Food and Nutrition Research; Toldrá, F., Ed.; Elsevier:

Cambrigde, UK, pp. 297–345.

- EC, 2006. Setting maximum levels for certain contaminants in foodstuffs. Official Journal of the European Union, Geneva.
- Richard, J.L., 2007. Some major mycotoxins and their mycotoxicosis—An overview. Int. J. Food Microbiol. 119, 3–10.
- Rodríguez, A., Rodríguez, M., Martín, A., Nuñez, F., Córdoba, J.J., 2012. Evaluation of hazard of aflatoxin B1, ochratoxin A and patulin production in dry-cured ham and early detection of producing mould by qPCR. Food Control 27, 118–126.
- Saad, S.M., Salem, R., Amin, R.A., Abu Zaid, K.E., 2015. The using of essential oils in improving mycological status of some meat products. BVMJ. 29, 85-96.
- Shaltout, F.A., Amin, R.A., Nassif, M.Z., Abd-Elwahab, S.A., 2014. Detection of aflatoxins in some meat products. BVMJ. 27, 368-374.
- Zadravec, M., Vahčić, N., Brnić, D., Markov, K., Frece, J., Beck, R., Lešić, T., Pleadin, J., 2020. A study of surface moulds and mycotoxins in Croatian traditional dry-cured meat products. Int. J. Food Microbiol. 317, 108459.